

# SEROLOGICAL SCREENING FOR TOXOPLASMOSIS IN DOMESTIC CATS OF CENTRAL KERALA

## Tresamol, P. V1\*., Arthi, A2., Merry Rincy, K3. and Gleeja, V. L.4

<sup>1</sup>Professor and Head,<sup>2</sup>MVSc scholar,<sup>3</sup> Research Assistant Department of Veterinary Epidemiology and Preventive Medicine <sup>4</sup>Associate Professor, Department of Statistics College of Veterinary and Animal Sciences, Mannuthy Kerala Veterinary and Animal Sciences University \*Corresponding author: tresamol@kvasu.ac.in

# ABSTRACT

The present study was designed to record the seropositivity rate of toxoplasmosis in domestic cats presented to the University Veterinary hospitals. Serum samples were collected from 43 cats and subjected to an indirect enzyme-linked immunosorbentassay(ELISA) for detection of IgG antibodies against Toxoplasma gondii using a commercially available kit. Out of 43 cats tested, 12 showed a positive reaction for antibodies against T. gondii (27.90 %). The rate of seropositivity was higher in cats above 3 years, male cats and those with outdoor access. But a statistically significant association was observed only between sex of cats and seropositivity rate whereas no significant association has been observed with age and rearing practice. The identification of seropositive cats in a community is important to estimate soil contamination with oocysts and determine populations with a high risk of exposure to T. gondii. The results of the present study stress on the need for establishing prevention strategies to minimize this neglected zoonosis.

**Keywords**: Domestic cats, *Toxoplasma* gondii. seropositivity ELISA

# **INTRODUCTION**

Toxoplasmosis caused by Toxoplasma gondii, is one of the most common parasitic zoonosis world-wide and infects a wide range of warm-blooded vertebrates, including humans. It is transmitted most commonly by the ingestion of tissue cysts in raw or undercooked meat. Infection also occurs by consumption of food or water contaminated with sporulated T. gondii oocysts. Cats are the definitive hosts and play an important role in the life cycle of T. gondii. They facilitate the genetic recombination between strains, as well as environmental contamination (Vilares et al., 2014). Cats can excrete millions of oocysts through faeces after primary infections. These oocysts are highly resistant in the environment, surviving for months in soil and water and after sporulation are infective for animals and humans (Dubey, 1995).

The identification of seropositive cats in a community is important to estimate the environmental contamination and determine the populations with a high risk of exposure to *T. gondii*. Since cats are one of the most popular pet animals, their infections pose zoonotic significance in the human population. The present study was designed to find out prevalence of *T.gondii* antibodies among domestic cats in Thrissur district of Kerala.

## MATERIALS AND METHODS

Domestic cats presented to the University Veterinary hospitals, Mannuthy and Kokkalai during the period of 2016-17 were screened for antibodies against *T. gondii*. Serum samples were collected from 43 cats and subjected to an indirect enzyme-linked immunosorbent assay (ELISA) for the detection of IgG antibodies against *T. gondii* (Multi-species ID Screen<sup>®</sup> Toxoplasmosis Indirect, IDVET, France).

Test serum samples  $(10\mu l)$  and positive and negative controls were diluted to 1:10 using 90  $\mu l$  of dilution buffer supplied with the kit. The diluted serum

samples were transferred to an ELISA microplate coated with P30 antigen of T. gondii and the plate was incubated at 21°C  $(\pm 5^{\circ}C)$  for 45 minutes. The wells were emptied and washed three times with 300 µl of 1X wash solution using an ELISA plate washer. Added hundred ul of 1X conjugate to each well and incubated at 21 °C (±5 °C) for 30 minutes. The wells were emptied and washing procedure was repeated as mentioned above. Hundred microlitre of substrate solution was added to each well. The plate was then incubated in the dark at 21 °C (±5 °C) for 15 minutes. Hundred microlitre of stop solution was added to each well in order to stop the reaction. Optical densities were measured at 450 nm using an ELISA plate reader (Varioskan Flash, Thermo Fisher Scientific) and SkanIt Software 2.4.5 RE. The test was considered valid after it satisfied the following conditions; The mean value of the positive control Optical density  $(O.D._{PC})$  was greater than 0.350 and the ratio of the mean O.D. values of the positive and negative controls is greater than three (O.D.<sub>PC</sub>/ O.D.<sub>NC</sub>> 3). For each sample, the seropositivity (S/P) percentage was calculated as follows: S/P per cent = (0.D.  $_{sample}$  - 0.D.  $_{NC}$  / 0.D.  $_{PC}$  - 0.D.  $_{NC}$ ) x 100. The result was considered as negative when S/P  $\% \leq 40$  per cent, Doubtful when 40 < S/P < 50 and positive when  $S/P \ge 50$ per cent.

#### **RESULTS AND DISCUSSION**

Out of the 43 serum samples tested, 12 were positive for antibodies against T. gondii and thus, seropositivity was 27.90%. The seropositivity ranged from 50 to 224% in positive cats. Varying levels of seroprevalence has been reported among cats from different parts of world (Zhang et al., 2010; Montazeri et al., 2020). Seroprevalence of T. gondii in domestic cats was estimated to be 30 to 40% worldwide (Dubev and beattie 2010). Montazeri et al. (2020) reported a higher seroprevalence among domestic cats in Australia (52%) and Africa (51%) and a lower prevalence in Asia (27%). Hatam-Nahavandi et al. (2021) also reported a lower prevalence in Asia (28.3%). The results of the present study are in agreement with these observations. A lower prevalence when compared to the present study was reported in serosurveys conducted at Bagkok Metropolitan region (6.5%) by Inpankaew *et al.* (2021) and the

Netherlands (18.2%) by Opsteegh *et al.* (2012). The variations in the seroprevalence of toxoplasmosis in different regions might be due to geographical factors or feeding and animal welfare conditions for cats in these areas.

The seropositivity of T.gondii in various age groups and sex of cats and under different rearing practice is depicted in Table1. The rate of seropositivity was higher in cats above three years (37.5%) and lowest in kittens below one year age (18.18%). But as per Fisher's exact test, no significant (p>0.05) association could be observed between age and prevalence of T.gondii antibodies. The higher seropositivity has been reported with increasing age of the cats ( Ahmad et al, 2014; Must et al., 2015) indicating that age is a risk factor for seropositivity in cats. Higher seropositivity in older cats might be related to increased exposure to T. gondii oocysts through food, water and outdoor activities (Xia et al., 2022).

Risk factor	Number tested	Number positive	Per cent
Age			
Below 1 year	11	2	18.18
1-3 years	24	7	29.17
Above 3 years	8	3	37.50
Sex			
Male	23	11	47.82**
Female	20	1	5.00
<b>Rearing practice</b>			
Indoor	15	3	20.00
Outdoor access	28	9	32.14
** Significant (p<0.01)			

 Table 1. Seroprevalence of Toxoplasmosis in domestic cats

The overall seropositivity of T. gondii infection was more in males (47.82%) than females (5%) and as per Fisher's exact test, there was significant (p<0.01) association between sex and seroprevalence of T.gondii. This result is in agreement with previous reports from Norway (Saevik et al., 2015). But no significant difference in the seroprevalence of T.gondii could be detected in male and female cats in Japan (Nogami et al., 1998), Brazil (Cardia et al., 2013), China (Xia et al., 2022) and Saudi Arabia (Mohammed et al., 2019). On the other hand, a higher seroprevalence of *T. gondii* has been reported in female cats in Hungary (Hornok *et al.*, 2008).

The seropositivity rate was higher in cats with outdoor access (32.14%) than those kept indoors (20%). But as per Fisher's exact test, there was no significant (p>0.05) association between rearing practice and seropositivity in the present study. Similar observations were also reported by Dubey et al. (2002) in rural Ohio and higher T. gondii seroprevalence found in cats with outdoor access has been attributed to their carnivorous behaviour and eating prey animals such as rodents and birds. Opsteegh et al. (2012) identified hunting behaviour and feeding of raw meat as the risk factors for higher prevalence of toxoplasmosis in stray cats, which could be

a potential target for intervention measures to prevent infections.

Currently there is an increase in the number of pet cats in Kerala. Since cats are one of the most popular pet animals, their infections may affect their owners and others living in their environment. The results of the present study stress on the need for establishing prevention strategies to minimize the occurrence of this neglected zoonosis in central Kerala. The proper disposal of cat litter, keeping cats indoors to minimize their acquisition of infection from prey or the environment, and reducing the feral cat population are the recommended measures. Also further research is warranted to assess the prevalence in other parts of the state and to investigate the links between cat ownership and human T. gondii infection.

## REFERENCES

- Ahmad, N. H., Irum, S. and Qayyum, M. 2014. Seroprevalence of IgG and IgM antibodies and associated risk factors for toxoplasmosis in cats and dogs from sub-tropical arid parts of Pakistan. *Trop. Biomed.* **31**(4): 777– 784.
- Cardia, D.F., Camossim, L.G., Neto, S., Langoni, H. and Bresciani, K.D. 2013. Prevalence of *Toxoplasma gondii* and

*Leishmania* spp. infection in cats from Brazil. *Vet. Parasitol.* **197**(3-4): 634–7. https://doi.org/10.1016/j. vetpar.2013.07.017.

- Dubey, J.P. 1995. Duration of immunity to shedding of *Toxoplasma gondii* oocysts by cats. *J. Parasitol.***81**:410-415
- Dubey, J.P. and Beattie, C. 2010 *Toxoplasmosis of animals and man.* 2nd ed. Boca Raton, Florida: CRC Press.
- Dubey, J. P., Saville, W. J. A., Stanek, J. F. and Reed, S. M. 2002. Prevalence of *Toxoplasma gondii* antibodies in domestic cats from rural Ohio. *J. Parasitol.* 88 (4): 802–803.
- Hatam-Nahavandi, K., Calero-Bernal, R., Rahimi, M.T., Pagheh, A.S., Zarean, M., Dezhkam, A. and Ahmadpour, E. 2021. *Toxoplasma gondii* infection in domestic and wild felids as public health concerns: a systematic review and meta-analysis. *Sci. Rep.* 11: 9509. https://doi.org/10.1038/s41598-021-89031-8
- Hornok, S., Edelhofer, R., Joachim, A., Farkas, R., Berta, K., Répási, A. and Lakatos, B. 2008. Seroprevalence of *Toxoplasma gondii* and *Neospora caninum* infection of cats in Hungary.

*Acta Vet. Hung.* **56** (1):81–8. https:// doi.org/10.1556/avet.56.2008.1.8.

- Inpankaew, T., Sattasathuchana, P., Kengradomkij, C. Thengchaisri, N. 2021. Prevalence of toxoplasmosis in semi-domesticated and pet cats within and around Bangkok, Thailand. *BMC Vet. Res.* **17**: 252.
- Mohammed, O.B., Omar, O.I., Elamin, E.A., Bushara, H.O., Omer, S.A., Alagaili, A.N.2019. Seroprevalence of *Toxoplasma gondii* in household and stray cats of Riyadh. Saudi Arabia *Vet. Ital.* 55(3):241–5.
- Montazeri, M., Mikaeili Galeh, T., Moosazadeh, Sarvi, S.,M.Dodangeh, S.,Javidnia, J., Sharif, M. and Daryani, A. 2020.The global serological prevalence of *Toxoplasma gondii* in felids during the last five decades (1967–2017): a systematic review and meta-analysis. *Parasites Vectors* 13, 82.
- Mus,tK., Lassen, B. and Jokelainen, P. 2015. Seroprevalence of and risk factors for *Toxoplasma gondii* infection in cats in Estonia. Vector-Borne Zoonotic Dis. 15(10), 597–601.
- Nogami, S., Moritomo, T., Kamata, H., Tamura, Y., Sakai, T., Nakagaki, K., et al.1998. Seroprevalence

against *toxoplasma gondii* in domiciled cats in Japan. *J. Vet. Med. Sci.* 60(9):1001–4. https://doi. org/10.1292/jvms.60.1001.

- Opsteegh, M., Haveman, R., Swart, A.N., Mensink-Beerepoot, M. E., Hofhuis, A., Langelaar M.F.M. and van der Giessen, J.W.B. 2012. Seroprevalence and risk factors for *Toxoplasma gondii* infection in domestic cats in The Netherlands, *Prev. Vet. Med*.104: 317-326
- Saevik, B.K., Krontveit, R.I., Eggen, K.P., Malmberg, N., Thoresen, S.I. and Prestrud, K. W. 2015. *Toxoplasma* gondii seroprevalence in pet cats in Norway and risk factors for seropositivity. J. Feline Med. Surg.

2015; 17 (12):1049–56. https://doi. org/10.1177/1098612X15569616.]

- Vilares, A., Gargate, M.J., Ferreira, I., Martins, S., Julio, C., Waap, H., Angelo, H. and Gomes, J.P. 2014. Isolation and molecular characterization of *Toxoplasma gondii* isolated from pigeons and stray cats in Lisbon, Portugal. *Vet. Parasitol.* 205(3–4), 506–511.
- Xia, N., Ji, N., Li, L.,Huang,Y.,Yang,C.,G uo,X.,Guo,Q.,Shen,B., Xiao,L. and Feng,Y. 2022. Seroprevalence and risk factors of *Toxoplasma gondii* in urban cats from China. *BMC Ve.t Res.* 18, 331. https://doi.org/10.1186/ s12917-022-03427-w